Processing of Slides Prior to Printing
SLIDE COATING PROTOCOL

(Make sure to wear powder free gloves at all times during these procedures!!)

1. Place "precleaned" Gold Seal microscope slides (Erie Scientific c#3011) into stainless steel slide rack, and place rack into glass tanks.

2. Prepare cleaning solution in a 2L beaker with a magnetic stir bar (enough for 6 30-slide tanks):
   - First add 840mL H_2O
   - Then add 1110mL 100% EtOH
   - Finally add 195g NaOH (this is an exothermic reaction so add SLOWLY)

3. Stir on magnetic stir plate until solution is clear.

4. Submerge rack in cleaning solution in glass tanks marked NaOH.

5. Shake at 60 rpm for 2 hr on orbital shaker.

6. Dump the cleaning solution and RINSE slides with DI H_2O by keeping the racks in the tanks marked NaOH. Cover the slides with DI H_2O and let sit for 3 min.

7. Repeat step 6 7-10 times.

8. Prepare poly L Lysine solution (make fresh ever time) by combining:
   - 1560mL H_2O
   - 195mL tissue culture PBS (no calcium and no magnesium)
     (GibcoBRL c# 10010-023)
   - 195mL poly L Lysine (Sigma c# P8920)

9. Place slide racks into glass tanks marked PLL and pour the solution made in step 8 into each tank covering the glass slides.

10. Shake 1 hr at 60 rpm on orbital shaker.

11. Place slide racks in glass tanks marked H_2O and pour DI H_2O into each tank covering the glass slides.

12. After 1 min of rinsing from step 11, remove the slide racks.

13. Centrifuge racks to remove free liquid (1,000 rpm for 5 min) in D-247.

14. Immediately transfer slides to clean slide box (do not use boxes with glued cork bottoms). Label the box with the coating date and your initials.

15. Allow slides to "age" for 2 weeks before printing.